

***In Vitro* protocol in brief test LymphoSign**

1. Reverse transcription

Reagents:

- Kit M-MLV RT
- dNTPs (10mM)
- Hexamers (100µM)
- ARN to be tested

Steps :

- Thaw the reagents
- Prepare reverse transcription mix
 - RT buffer (1,25 µL)
 - DTT (0,5 µL)
 - dNTPs (1 µL)
 - Hexamers (1 µL)
- Deliver 3,75 µL mix per tube
- Add 2 µL of RNA sample
- Vortex and centrifuge
- Put samples in thermocycler and start program 1 step 1a
- Add 0,5 µL of M-MLV RT
- Centrifuge
- Put samples in thermocycler and start program 1 step 1b
- Once the program is over and the block cool down at 4°C, put out tubes
- Put tubes on ice or cooling block



Then proceed to step 2 or store cDNA products between -30°C et -15°C.

2. Hybridization of probes

Reagents:

- SALSA MLPA Buffer
- GEP- LSPM** probe Mix

- Thaw reagents
- Prepare Hybridization mix
 - Salsa MLPA Buffer (1,5 µL)
 - GEP-LSPM** Probe Mix (1,5 µL)
- Vortex and centrifuge
- Deliver 3 µL of mix per cDNA tubes
- Centrifuge
- Put samples in thermocycler and start program 1 step 2

3. Ligation

Reagents:

- | | |
|--|---|
| <ul style="list-style-type: none"> <input type="checkbox"/> SALSA Ligase Buffer A <input type="checkbox"/> SALSA Ligase Buffer B <input type="checkbox"/> SALSA Ligase 65 <input type="checkbox"/> Nuclease free water | <ul style="list-style-type: none"> <input type="checkbox"/> Thaw reagents <input type="checkbox"/> Prepare ligation mix <ul style="list-style-type: none"> <input type="checkbox"/> Nuclease free water (25 µL) <input type="checkbox"/> Salsa Ligase Buffer A (3 µL) <input type="checkbox"/> Salsa Ligase Buffer B (3 µL) <input type="checkbox"/> Vortex and centrifuge <ul style="list-style-type: none"> <input type="checkbox"/> Salsa Ligase 65 (1 µL) <input type="checkbox"/> After 1h of hybridization, deliver 32 µL of mix per cDNA tubes <input type="checkbox"/> Start program 1 étape 3 |
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Then proceed directly to step 4 or store the ligation products between -30°C and -15°C.



After this step, do not store the products at high temperature (e.g. 4°C or at room temperature) in order to avoid non-specific ligations which could result from a residual activity of the enzyme.

4. Amplification and incorporation of barcodes and adaptors

Reagents:

- | | |
|--|---|
| <ul style="list-style-type: none"> <input type="checkbox"/> GEP-BC-xxx
Barcodes <input type="checkbox"/> Q5 MasterMix <input type="checkbox"/> Nuclease free water | <ul style="list-style-type: none"> <input type="checkbox"/> Thaw the reagents <input type="checkbox"/> Prepare PCR mix <ul style="list-style-type: none"> <input type="checkbox"/> Q5 MasterMix (12,5 µL) <input type="checkbox"/> Nuclease free water (5,5 µL) <input type="checkbox"/> Vortex and centrifuge <input type="checkbox"/> Deliver 18 µL of mix per wells of PCR plate <input type="checkbox"/> Add 5 µL of ligation product in each well <input type="checkbox"/> Add 2 µL of barcode (GEP-BC-xxx) in each well |
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Use different barcodes BEP-BC-xxx for each tested samples.

- Put samples in thermocycler and start program 2



Then proceed directly to step 5 or store the PCR products between -30°C and -15°C.



Do not store these products for prolonged periods at high temperature (e.g. 4°C in the thermocycler or at room temperature).

5. Purification et dosage sequencing librairies

Reagents:

- 100% Ethanol
- Nuclease free water
- AMPure XP Magnetic beads
- TE Buffer
- Qubit dsDNA HS Assay

Steps :



Ensure beads are completely re-suspended before use.

- Purify 25 μ L products with 45 μ L AMPure XP
- Elute PCR products in 50 μ L of TE buffer



After purification, librairies can be stored between -30°C et -15°C before sequencing.

- Dose 10 μ L of each library by fluorimetry



This rapid protocol completes the notice. It does not dispense with complete reading of the notice.